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ON THE WAY TO LIPOSIDOMYCINS, NEW NUCLEOSIDE ANTIBIOTICS. ACCESS TO THE HOMOCHIRAL DIAZEPANONE CORE

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ABSTRACT

Access to the homochiral diazepanone core of liposidomycins has been canied out through the regiospecific nucleophilic opening of an enantiomerically pure α -amino- β , γ epoxy-acid precursor, by an L-amino acid derivative, on one hand and cyclisation by a peptidic coupling reaction, on the other hand.

INTRODUCTION

The ribosyl-diazepanone moiety is a common component in liposidomycins, a new class of nucleoside antibiotics, isolated from the culture filtrate and mycelia of *Streptomyces griseosporeus*,¹ which strongly inhibit bacterial peptidoglycan synthesis.² At least twelve active components were obtained and the structures of the three major constituents, namely liposidomycins A, B and C were previously described and partially elucidated3 (Scheme 1). These compounds contain 5'-substituted uridine, *5* **amino-5-deoxyribose-2-sulfate** and 1,6diazepan-Zone moieties and differ essentially in the structure of the lipid side-chain. However, the absolute and relative configurations at C_2 , C₅ and C₆ are still unknown.

RESULTS AND DISCUSSION

When we began our study, only two syntheses of the 1,4-diazepan-2-one system had been reported⁴ and both of them required separation of stereo- or regioisomers; we have been aiming at developing an approach **using** homochiral precursors. The two proposed key steps (Scheme 1) are: the regiospecific nucleophilic opening of **an** epoxide by the amino group of an α -ribosyl amino acid and cyclisation by a peptidic coupling reaction.5 More recently, reports concerning a similar approach, but involving a glycine derivative, 6.7 prompted us to submit our results in extenso.

Taking the hypothesis that the biosynthetic route to liposidomycins involves naturally occurring amino acids, we postulated that the absolute configuration at C2 and *C5* was *S*. For this reason, we chose to study the nucleophilic opening of a β , γ -epoxy-Lrhreo-amino acid derivative by L-phenylalanine (Scheme 2), as a test of the feasibility of our synthetic scheme.

Scheme 3: a) Tf₂O, 2,6-lutidine, -78 °C. b) MeNH₂, EtOH, 70%. c) LiAlH₄, THF, Δ . d) **TBDPSCI, DMF,** imidazole, **86%.** e) BnBr, K2CO3, **DMF, 88%. f)** PhCH20COC1, K2CO3, **DMF,** 90%. g) **TFA,** H20,95%. h) Ph3P, DIAD, 67% for **7a** and 75% for **7b.**

D-Isoascorbic acid was revealed to be a **good** precursor of the epoxy amino acid derivative. In fact, due to stability reasons, we first synthesized a **C4** building block in which the carboxylic acid function was reduced to a primary alcohol and the amine function was protected as its N-benzyl or N-benzyloxycarbonyl derivative (Scheme **3).**

Ethyl 3,4-0-methylethylidene D-erythronate **1** was obtained according to a known route.8 Activation of the secondary alcohol by reaction with trifluoromethane sulfonic anhydride in the presence of 2,6-lutidine in CH_2Cl_2 at -78 °C was followed by nucleophilic substitution of the triflate with an excess of methylamine in ethanol at 20 °C. This cleanly afforded the secondary amine **2** in 70% overall yield from **1.** LiAW reduction of the ester function in refluxing THF gave the primary alcohol **3** (quantitative yield) which was then protected as **its O-TBDPS** derivative in **86%** yield. The secondary amine was subsequently

Scheme 4: a) tBuOH, NaH, Δ, 75%. b) HCO₂NH₄, MeOH, 10% Pd/C, 84%. c) TFA, H₂O *65%.* d) DCC, DMAP, DMAP.HCI, CHCI,, **A** then TBDPSCI, imidazole, DMF, **45** OC.

protected either as its N-benzyl (Sa) or N-benzyloxycarbonyl **(Sb)** derivative in good yield. Acidic hydrolysis of the acetonide moiety afforded the corresponding **diols 6a and 6b** in 95% and **92%** yield, respectively. Epoxide formation cleanly occurred under Mitsunobu conditions to give the expected epoxides 7a and **7b** in *67%* and 75% yield, respectively.

Although many examples of nucleophilic opening of an epoxide by amines have been described,⁹ relatively few involve the opening by an α -amino acid.¹⁰ The nucleophilic opening of the epoxide $7a^{11}$ by reaction with *tert*-butyl L-phenylalaninate hydrochloride could be efficiently achieved (Scheme 4) in the presence of sodium *tert*butoxide in tert-butyl alcohol at 100 $^{\circ}$ C for 24 hours. The expected product 8¹² was obtained in this way in **75%** yield.13 Hydrogenolysis of the N-benzyl moiety cleanly occurred in methanol in the presence of **an** excess of ammonium formate and Pd/C (10%) and afforded 9 in 84% yield.¹⁴ Both silyl ether and *tert*-butyl ester deprotections were then effected by acid hydrolysis (TFA, H20, 65% crude yield). **A** sample of the resulting salt **10** was successfully cyclized in the presence of an excess of DCC, DMAP, DMAP.HCI in refluxing chloroform.¹⁵ In order to decrease the polarity of the product and facilitate chromatographic purification, the crude mixture was persilylated (TBDPSC1 excess, imidazole, DMF) to afford the diazepanone 11 in moderate yield.¹⁶

The 1,4-diazepan-2-one structure was unequivocally established¹² by mass spectroscopy and 2D NMR experiments [COSY ¹H-¹H, COSY ¹H-¹³C, and long range $1H-13C$ correlation (PFG-HMBC)¹⁷].

CONCLUSION

We have shown here that the diazepanone core of liposidomycins can be obtained according to the synthetic route we proposed, namely: the regiospecific nucleophilic opening of an epoxide by the amino group of an α -ribosyl amino acid and cyclisation by a peptidic coupling reaction. Our current efforts are directed towards obtaining the ribosyldiazepanone present in the liposidomycins structure.

EXPERIMENTAL

¹H NMR (250 MHz) and ¹³C NMR (63 MHz) spectra were recorded in CDCl3 (unless otherwise indicated). High Resolution Mass Spectra were recorded in Service de Spectrométrie de Masse, Université Pierre et Marie Curie. Specific rotations were measured on a Perkin Elmer 241C polarimeter with sodium (589 nm) or mercury (365 nm) lamps. All reactions were carried out under a nitrogen atmosphere using dried solvents and were monitored by thin-layer chromatography with Merck 6OF-254 precoated silica (0.2 mm) on glass. Flash chromatography was performed with Merck Kieselgel 60H (5-40 μ m). Spectroscopic (¹H and ¹³C NMR, MS) and/or analytical data were obtained using chromatographically homogeneous samples.

Ethyl (2S, 3s) -2-N- Me t h y I am i no -3,4- *0* **-met h y le t h y I i den e-3,4 dihydroxybutanoate (2).** To a solution of the hydroxy ester l(4.6 *g,* 22.5 mmol) in CH₂Cl₂ (63 mL), at -78^oC, were successively added 2,6-lutidine (3,67 mL, 1.4 eq) and trifluoromethanesulfonic anhydride (5.33 mL, 1.4 eq). After 30 min, thin-layer chromatographic analysis (Et0Ac:cyclohexane 6:4) of the reaction mixture revealed the absence of starting material $(Rf 0.26)$ and the presence of the expected triflate $(Rf 0.82)$. Methylamine (11.2 mL, 4 eq) was then added and the mixture was stirred at 20 $^{\circ}$ C for 12 h. Concentration *in vucuo* and purification by flash chromatography (EtOAc: cyclohexane:Et3N 6:4:0.01, pre-saturation of the column with 1% Et3N, $Rf(0.34)$ afforded 3.4 g (70%) of the amino ester 2: $[\alpha]_D$ -40 (c 1.00, CH₂Cl₂); ¹H NMR δ 1.28 (t, 3H, J = 7 *Hz,* OEt), 1.41, 1.33 (2s, 6H, CMe2), 2.42(s, 3H, NHMe), 3.18 (d, **lH,** J2,3 = 6 Hz, **Hz,** H-47, 4.22 (9, **2H,** *J* = 7Hz, OEt), 4.28 (m, lH, H-3); l3C NMR *6* 14.3 (OEt), 25.2, 26.4 (CMe2), 34.8 (NMe), 61.1 (OEt), 65.1 (C-2), 66.5 (C-4), 75.9 (C-3), 109.7 (CMe₂), 172.0 (C-1); IR (neat) : 3400-3300, 1740 cm⁻¹. H-2), 3.92 (dd, lH, *J4,4'* = -8 Hz, *J4.3* = 7 Hz, H-4), 4.01 (dd, lH, **54',4** = -8,54*,3 = 7

Anal. Calcd for C10Hig04N: C, *55.28;* H, 8.81; N, 6.45. Found: C, 55.31; H, 8.82; N, 6.45.

(2S,3R)-3-N-Methylamino-1,2-O-methylethylidene-butan-1,2,4-triol **(3).** To a suspension of lithium aluminium hydride (854 mg, 1.44 eq) in THF (15.8 mL), at 0 "C, was dropwise added a solution of the amino ester **2** (3.39 g, 15.6 mmol) in THF (10.5 mL). After 1.5 h stirring at 20 °C and 1.5 h at 65 °C, the mixture was cooled to 0 °C. Careful additions of water (854 μ L), of a 15% aqueous solution of NaOH (854 μ L) and finally of water (2.6 **mL)** led to the formation of a white precipitate which was filtered off and successively rinsed with ether and boiling chloroform. The organic extracts were then dried (Na2S04) and concentrated *in vacuo to* afford 2.68 g (98%) of the crude alcohol **3** which was used in the next step without further purification. A sample was purified by flash chromatography (EtOAc:cyclohexane:Et3N 7:3:0.01; pre-saturation of the column with 1% Et3N). 'H NMR **6** 1.32,1.39 (2s, 6H, CMe2), 2.42 (s, 3H, NHMe), 2.50 (ddd, IH, $J_{3,2} = 7.5$ Hz, $J_{3,4} = 3$ Hz, $J_{3,4} = 4$ Hz, H-3), 3.33 (dd, 1H, $J_{4,4} = -11.5$ Hz, $J_{4,3} =$ 3 Hz, H-4), 3.68 (dd, 1H, $J_{4,4} = -11.5$ Hz, $J_{4,3} = 4$ Hz, H-4'), 3.73 (dd, 1H, $J_{1,1} = -8$ Hz, $J_{1,2} = 6$ Hz, H-2), 4.04 (dd, 1H, $J_{1',1} = -8$ Hz, $J_{1',2} = 6$ Hz, H-1'), 4.15 (ddd, 1H, $J_{2,3} = 7.5$ Hz, $J_{2,1} = J_{2,1} = 6$ Hz, H-2).

(2R,3S)- **l-tert-Butyldiphenylsilyloxy- 2-N-methylamino- 3,4-0 methylethylidene-butan-3,4-dioI (4).** To a solution of the amino alcohol **3** (2.63 g, 15 mmol) in DMF (91.5 mL) were added imidazole (2.24 g, 2.2 eq) and *tert***butyldiphenylsilylchloride** (4.3 **mL,** 1.1 eq). The resulting mixture was stirred for 48 h **at** 20 "C and concentrated *in vacuo.* Flash chromatography of the residue (cyclohexane:EtOAc:Et3N 1:1:0.01; pre-saturation of the column with 1% Et3N, Rf0.31) gave 5.35 g (86%) of the silyl ether 4 as a white solid: mp 56 °C; $[\alpha]_D$ -9 (c 0.94, CH2C12); lH NMR *6* 1.04 **(s,** 9H, CMe3), 1.33, 1.36 (2s, 6H, CMe2), 2.34 *(s,* 3H, NHMe), 2.55 (m, 1H, H-2), 3.57 (dd, 1H, $J_{1,1'} = -10.5$ Hz, $J_{1,2} = 6$ Hz, H-1), 3.64 (dd, 1H, $J_{1',1}$ = -10.5 Hz, $J_{1',2}$ = 4.5 Hz, H-1'), 3.78 (dd, 1H, $J_{4,4'}$ = -7.5 Hz, $J_{4,3}$ = 7.5 Hz, H-4), 4.01 (dd, 1H, $J_{4',4} = -7.5$ Hz, $J_{4',3} = 6$ Hz, H-4'), 4.11 (ddd, 1H, $J_{3,4} = 7.5$ Hz, $J_{3,2} = 7$ Hz, $J_{3,4'} = 6$ Hz, H-3), 7.39, 7.62 (2m, 10H, Ph); ¹³C NMR δ 19.1 (CMe₃), **25.4** (CMe2), 26.8 (CMe3), 34.7 (NMe), 62.9 (C-l), 63.5 (C-2), 67.1 (C-4), 77.3 (C-3), 108.4 (CMe2), 127.7, 129.7, 133.1, 135.5 (Ph).

8.56; N, 3.53. Anal. Calcd for C24H3503NSi: C, 69.69; H, 8.53; N, 3.39. Found: C, 69.59; H,

(2R, 3S)-1-tert-Butyldiphenylsilyloxy-2-N-methylbenzylamino-3,4-**O-methylethylidene-butan-3,4-diol (5a).** To a solution of the amine **4** (550 mg, 1.3 mmol) in DMF (6.5 **mL)** were successively added potassium carbonate (276 mg, 1.5 eq) and benzyl bromide (217 μ L, 1.37 eq) after cooling to 0 °C. The mixture was then stirred at 20 \degree C for 12 h. After having discarded the excess of potassium carbonate by

filtration, the filtrate was concentrated *in* vacuo. To the resulting residue, ether was added and the insoluble potassium chloride was filtered off. The organic layer was then dried $(Na₂SO₄)$ and concentrated *in vacuo*. Flash chromatography of the residue (cyclohexane:EtOAc:Et3N 9:1:0.01, pre-saturation of the column with 1% Et3N, $Rf(0.38)$) afforded 590 mg (88%) of the benzylated amine 5a: $\lbrack \alpha \rbrack_D$ -4 $\lbrack c \rbrack_1$, $\lbrack 0$, CH₂Cl₂), $\lbrack \alpha \rbrack_{365}$ -14 (c1.00, CH2C12); IH NMR 6 1.04 **(s,** 9H, CMe3), 1.35, 1.36 (2s, 6H, CMe2), 2.36 (s, 3H, NMe), 2.79 (ddd, 1H, J2.3 = *6.5* Hz, J2.1 = 6 Hz, J2,1~ = *5.5* Hz, H-2), 3.72 (dd, 1H, $J_{1,1'} = -10.5$ Hz, $J_{1,2} = 6$ Hz, H-1), 3.76 (m, 1H, H-4), 3.83 (dd, 1H, $J_{1',1} = -10.5$ Hz, J11.2 = 5.5 *Hz,* H-l'), 3.68, 3.93 (AB, 2H, *JA,B* = -13.5 Hz, CH2Ph), 3.97 (dd, lH, $J_{4,4} = -8$ Hz, $J_{4,3} = 6$ Hz, H-4'), 4.33 (ddd, 1H, $J_{3,4} = 8$ Hz, $J_{3,2} = 6.5$ Hz, $J_{3,4} = 6$ Hz, H-3), 7.26, 7.40, 7.65 (3m, 15H, Ph); *3C NMR *6* 19.1 (CMe3), 25.6 (CMe2). 26.6, 26.9 (CMe3). 38.8 (NMe), 60.2, 62.5 (C-1, NCHzPh), 64.6 (C-2), 67.5 (C-4), 76.0 (C-3), 108.4 (CMe2), 126.6, 127.7, 128.1, 128.7, 129.7, 133.2, 135.6, 140.4 (Ph).

Anal. Calcd for C31H4103NSi: C, 73.91; H, 8.20; N, 2.78. Found: C, 73.96; H, 8.29; N, 2.77.

(2R,3S)-l-tert-Butyldip **henylsilyloxy-2-N-methyl** benzyloxycarbonyl**amino-3,4-O-methylethylidene-butan-3,4-diol** (5b). To a solution of the amine **4** (230 mg, 0.56 mrnol) in DMF (2.7 **mL)** were successively added potassium carbonate (93 mg, 1.3 eq) and benzylchloroformate (87.5 μ L, 1.1 eq) after cooling to 0 °C. After 30 min stirring at 20 \degree C, further addition of both potassium carbonate (0.6 eq) and benzylchloroformate (0.55 eq) allowed completion of the reaction within 30 min stirring at 20 **OC.** The mixture was then filtered and washed with DMF and the filtrate was concentrated *in vucuo.* The resulting residue was dissolved in chloroform and washed with brine. The organic layer was then dried (Na₂SO₄) and concentrated *in vacuo*. Flash chromatography (cyclohexane:EtOAc 8:2, Rf 0.38) afforded 276 mg (90%) of the carbamate 5b: 1H NMR 6 1.00 **(s,** 9H, CMe3), 1.27, 1.29 (2s, 6H, CMe2), 2.96 (s, 3H, NMe), 3.49-3.97 (m, 4H, H-1,1',4,4'), 4.13, 4.28 (2m. 2H, H-2,3), 5.10 (m, 2H, OCHzPh), 7.35, 7.64 (2m, 15H, Ph).

3,4-diol (6a). To the acetonide 5a (660 mg, 1.3 mmol) in water (9.4 mL) at 0° C was dropwise added trifluoroacetic acid (9.4 mL). The mixture was stirred for 2 h and concentrated *in vacuo.* The resulting residue was dissolved in MeOH and silica gel was then added. After concentration *in vacuo,* the crude, absorbed on silica gel, was purified by flash chromatography (EtOAc:cyclohexane:Et3N 6:4:0.01, pre-saturation of the column with 1% Et₃N, Rf 0.24) to give 625 mg (95%) of the diol 6a: $[\alpha]_D$ -6 *(c* 1.20, CH₂Cl₂), **(2R,3S)-l-tert-Butyldiphenylsilyloxy-2-N-met** hyl benzylaminobutan-

[a1365 -20 (c 1.20, CH2C12); 1H NMR **6** 1.07 **(s,** 9H, CMe3), 2.22 **(s,** 3H, NMe), 2.84 (ddd, 1H, $J_{2,3} = 9$ Hz, $J_{2,1'} = 6$ Hz, $J_{2,1} = 5$ Hz, H-2), 3.44 (dd, 1H, $J_{4,4'} = -11.5$ Hz, $J_{4,3} = 4$ Hz, H-4), 3.60 (ddd, 1H, $J_{3,2} = 9$ Hz, $J_{3,4} = 4$ Hz, $J_{3,4'} = 3.5$ Hz, H-3), 3.74 Hz, H-1), 3.58, 3.81 (AB, 2H, $J_{A,B}$ = -13 Hz, CH₂Ph), 3.87 (dd, 1H, $J_{1,1}$ = -11.5 Hz, J1u.2 = 6 Hz, H-l'), 7.27, 7.44, 7.66 (3m, 15H, Ph); 13C NMR **6** 19.0, 26.8 (CMe3), 37.0 (NMe), 59.5, 60.1 (C-4, NCH2Ph), 63.9 (C-I), 65.2 (C-2), 68.4 (C-3), 127.1, 127.6, 128.3, 128.7, 129.8, 129.9, 132.7, 135.5, 135.6, 138.8 (Ph); HRMS calcd for C28H3703NSi: 463.2543. Found: 463.2543. (dd, 1H, $J_{4,4} = -11.5$ Hz, $J_{4,3} = 3.5$ Hz, H-4'), 3.77 (dd, 1H, $J_{1,1} = -11.5$ Hz, $J_{1,2} = 5$

(2R,3S)-l-tert-Butyldiphenylsilyloxy-2-N-met hylbenzyloxycarbonylaminobutan-3.4-diol (6b). To the acetonide 5b $(194 \text{ mg}, 0.36 \text{ mmol})$, at 20 °C, was added an aqueous solution of acetic acid $(5.9 \text{ mL}, \text{AcOH:}H_2O 6:1)$. After 48 h stirring at 20 °C, the mixture was concentrated *in vacuo* and purified by flash chromatography (EtOAc:cyclohexane 7:3, Rf 0.31) to afford 165 mg (92%) of the diol 6b: $[\alpha]_D +8$ (c 1.03, CH2CI2); IH NMR **6** 1.02 **(s,** 9H, CMe3), 3.00 **(s,** 3H, NMe), 3.49 (m, 2H, H-**4,4'),** 3.86 (m, 2H, H-l,l'), 4.03 (m, 2H, H-2,3), 5.13 (m, 2H, OCH2Ph), 7.34, 7.61 (2m, 15H, Ph); 13C **NMR 6** 19.1, 26.8 (CMe3), 33.7 (NMe), 60.1 (C-2), 62.4, 63.9 (C- (Ph), 158.2 (CO₂CH₂Ph); HRMS calcd for C₂₉H₃₇O₅NSi (M⁺-tBu): 450.1737. Found: 450.1733. 1,4), 67.5 (OCH2Ph), 71.7 (C-3), 127.8, 128.1, 128.5, 129.9, 132.9, 135.5, 136.5

(2R,3S)- 1-tert-Butyldiphenylsilyloxy-2-N-methylbenzylamino-3,4**epoxy-butane (7a).** To a solution of triphenylphosphine (2.32 **g,** 1.32 eq) in toluene (31 mL) at 0 °C, was dropwise added diisopropyl azodicarboxylate (1.74 mL, 1.32eq). After 15 min stirring at 0° C, a solution of the diol 6a $(3.1 \text{ g}, 6.7 \text{ mmol})$ -which was previously twice diluted with toluene (10 mL) and concentrated *in vucuo* in order to avoid any trace of water- in toluene (10 **mL)** was added and the resulting mixture was stirred at 0 "C for 1 h. After concentration *in vucuo,* the residue was gradually heated to 130 "C under *vucuo* (0.2 mm Hg) for **90** min. An intermediate trap cooled in *dry* ice was used in order to condense any volatile materials. The crude product was then purified by flash chromatography (cyclohexane:EtOAc:Et₃N 9:1:0.01, pre-saturation of the column with 1% Et₃N, *Rf* 0.26) to give 2 g (67%) of the expected epoxide 7a: $[\alpha]_D$ -10 (c 1.00, CH₂Cl₂); 'H NMR **6** 1.04 **(s,** 9H, CMe3), 2.30 **(s,** 3H, NMe), 2.41 (ddd, lH, J2.3 = 7.5 **Hz,** J2.1 (dd, 1H, $J_{4,4} = -5$ Hz, $J_{4,3} = 4$ Hz, H-4'), 3.13 (ddd, 1H, $J_{3,2} = 7.5$ Hz, $J_{3,4'} = 4$ Hz, $J_{3,4} = 2.5$ Hz, H-3), 3.75 (s, 2H, CH₂Ph), 3.83 (AB from ABX, 2H, $J_{A,B} = -10.5$ Hz, *JA,X* = 5.5 Hz, *JB,X* = 6.5 Hz, H-l',l), 7.28, 7.40, 7.63 (3m, 15H, Ph); 13C NMR 6 $= 6.5$ Hz, $J_{2,1'} = 5.5$ Hz, H-2), 2.58 (dd, 1H, $J_{4,4'} = -5$ Hz, $J_{4,3} = 2.5$ Hz, H-4), 2.74

19.1, 26.8 (CMe3), 38.8 (NMe), 44.8 (C-4), 51.6 (C-3), 59.8 (CH2Ph), 63.3 (C-1), 66.7 (C-2), 126.7, 127.7, 128.1, 128.6, 129.7, 133.3, 135.6, 139.9 (Ph).

Anal. Calcd for C₂₈H₃₅O₂NSi: C, 75.46; H, 7.92; N, 3.14. Found: C, 75.41; H, 7.93; N, 3.16.

(2R,3S)-l-tert-Butyldiphenylsilyloxy-2-N-met hyl benzyloxycarbonylamino-3,4-epoxy-butane (7b). A solution **of** the diol **6b** (57 mg, 0.11 mmol) in toluene (1 mL) was twice concentrated *in vucuo* in order to avoid any trace of water. A solution of mphenylphosphine (38 mg, 1.3 eq) in toluene (1 **mL)** was then added and the mixture was cooled to 0° C prior to the addition of diisopropyl azodicarboxylate (29 μ L, 1.3 eq). After 30 min stirring at 0 °C, the mixture was concentrated *in vacuo* and gradually heated to 130 °C under *vacuum* (0.02 mm Hg) for 2 h. Flash chromatography of the crude product (cyclohexane:EtOAc 8:2, $Rf(0.27)$ afforded 41 mg (75%) of the epoxide 7b: $[\alpha]_D$ -3 **(c** 1.00, CH2C12); 1H NMR 6 1.02 **(s,** 9H, CMe3). 2.57 (m, IH, H-4), 2.75 (m, lH, H-47, 2.94 **(s,** 3H, NMe), 3.08-3.17 (m, lH, H-3), 3.80-3.87 (m, 2H, H-l,l'), 4.06 (m, IH, H-2), 5.11 (m, *W,* OCHzPh), 7.35, 7.61 (2m, **15H,** Ph); 13C NMR 6 19.1, 26.7 (CMe3), 31.7 (NMe), 45.7 (C-4), 50.7 (C-3), 59.2 (C-2), 62.2 (C-l), 67.2 (OCH₂Ph), 127.8, 127.9, 128.4, 129.8, 133.0, 135.5, 136.7 (Ph), 156.6 (CO₂CH₂Ph).

tert-Butyl N-[(2R,3R)-4-tert-butyldiphenyIsilyloxy-2-hydroxy-3-(Nmethylbenzylamino)butyl]-S-phenylalaninate (8). To a solution of sodium *tert*butoxide, prepared by the addition of *tert*-butyl alcohol (2 mL) to sodium hydride (41 mg) , 1.9 eq), **was** added rerr-butyl L-phenylalaninate hydrochloride (463 **mg,** 2 eq). After stirring for 30 min at 20 °C, a solution of the epoxide 7a (400 mg, 0.9 mmol) in *tert*-butyl alcohol (2.5 mL) was added and the mixture was then heated to 100 °C for 24 h. After concentration *in vucuo* the residue was added with chloroform and filtered. The filtrate was dried (Na₂SO₄) and concentrated *in vacuo*. Flash chromatography of the resulting oily residue (toluene: MeOH:Et3N 98:2:0.01, pre-saturation of the column with 1% Et3N, Rf 0.32) afforded 450 mg (75%) of the expected product 8: $[\alpha]_D$ +5 $(c \ 1.25, \text{CH}_2\text{Cl}_2)$, $[\alpha]_{365}$ +27 (c 1.25, CH₂Cl₂); ¹H NMR δ 1.10 (s, 9H, SitBu), 1.34 (s, 9H, CO₂tBu), 2.28 (s, 3H, NMe), 2.32 (dd, 1H, $J_{1,1'} = -12.2$ Hz, $J_{1,2} = 5.4$ Hz, H-1), 2.82-2.85 (m, 3H, H-1', CH₂Ph), 2.94 (X from ABX, 1H, $J_{X,A} = 9$ Hz, $J_{X,B} = 5.4$ Hz, H-3), 3.24 (t, Hz, H-2), 3.77 (AB from ABX, 2H, *JA,B* = -5.4 Hz, *JA,X* = 9 Hz, **JB,X** = 5.4 Hz, H-20H, Ph); ¹³C NMR δ 19.1, 26.9 (SitBu), 28.0 (CO₂tBu), 37.1 (NMe), 39.8 (CH₂Ph), 50.4 (C-I), 59.8, 60.5 (C-4, NCH2Ph), 63.8, 66.2, 67.3 (C-2,3,Phe), 80.8 (C02tBu), 126.3, 127.1, 127.8, 128.1, 128.3, 128.8, 129.4, 129.8, 129.9, 133.1, 135.6, 135.7, 137.9, 139.5 (Ph), 173.7(CO2tBu). 1H, $J_{HPhe,CH2Ph} = 7.1$ Hz, H-Phe), 3.57 (ddd, 1H, $J_{2,3} = 9$ Hz, $J_{2,1} = 5.4$ Hz, $J_{2,1} = 3$ 4,4'), 3.69, 3.92 (2d, 2H, JNCH2ph = -13.2 Hz, NCH2Ph), 7.11-7.30, 7.40, 7.66 (3m,

Anal. Calcd for C41H5404NzSi: C, 73.83; H, 8.16; N, 4.2. Found: C, 73.81; H, 8.34; N, **4.1.**

tert-Butyl N-[(2R,3R)-4-tert-butyldiphenylsilyloxy-2-hydroxy-3-(N**methylamino)butyl]-S-phenylalaninate (9).** To a solution of the benzylated amine **8** (290 mg, 0.44 mmol) in MeOH (2.9 **mL)** at 20 **"C,** were successively added 10% palladium on charcoal (290 mg) and ammonium formate (110 mg, 4 eq). After 3.5 h stirring at 20 $^{\circ}$ C, the catalyst was removed by filtration through a celite pad and rinsed with chloroform previously filtered on potassium carbonate. Concentration *in vucuo* of the filtrate followed by a flash chromatography (toluene: $MeOH:Et₃N$ 85:15:0.01, presaturation of the column with 1% Et₃N, $Rf(0.30)$ afforded 211 mg (84%) of 9: α _D -3 (c 1.04, pyridine), $[\alpha]_{365}$ -4 (c 1.04, pyridine); ¹H NMR δ 1.11 (s, 9H, SitBu), 1.38 (s, 9H, COztBu), 2.42 **(s,** 3H, NMe), 2.92 (m, 2H, H-1,3), 3.04 (m, 2H, CH2Ph), 3.28 (dd, 1H, $J_{1,1} = -11.5$ Hz, $J_{1,2} = 4$ Hz, H-1'), 3.66 (t, 1H, $J_{HPhe,CH2Ph} = 7$ Hz, H-Phe), 3.93 (dd, 1H, $J_{4,4'} = -10$ Hz, $J_{4,3} = 4.5$ Hz, H-4), 4.04 (dd, 1H, $J_{4',4} = -10$ Hz, $J_{4',3} = 5$ Hz, H-4'), 4.19 (dt, 1H, $J_{2,3} = J_{2,1} = 5$ Hz, $J_{2,1'} = 4$ Hz, H-2), 7.27, 7.47, 7.85 (3m, 15H, Ph); ¹³C NMR δ 19.4, 27.0 (SitBu), 27.9 (CO₂tBu), 35.1 (NMe), 40.0 (CH₂Ph), 51.4 (C-l), 62.9 (C-4), 64.2, 64.3 (C-3,Phe), 70.1 (C-2), 80.5 (COztBu), 126.6, 128.2, 128.4, 129.8, 130.1, 134.0, 136.0, 138.6 (Ph), 174.0 (CO2tBu).

Anal. Calcd for C34H48O4N2Si: C, 70.79; H, 8.39; N, 4.86. Found: C, 70.97; H, 8.46; N, 4.69.

 $N-[2R,3R)-2,4-dihydroxy-3-(N-methylamino)butyl]-S-phenvlalanine$ **(10).** The ester **9** (210 mg, 0.37 mmol) in a 1:l (v/v) aqueous solution of trifluoroacetic acid **(4** mL) was stirred at 80 "C for 3 h. TLC monitoring of the mixture (toluene:MeOH:Et3N 85: 15:O.Ol) revealed the disappearance of starting material *(Rf0.30)* and the formation of the ammonium bis-trifluoroacetate salt *(RfO).* Concentration *in vucuo* afforded 120 mg (65%) of the crude salt 10: ¹H NMR (pyridine-d5) δ 3.44-3.97 (3m, **5H,** H-1,1',3, CHZPh), 4.30 (m, 2H, H-4,4), 4.54 (m, lH, H-Phe), 4.87 (m, lH, H-2), 7.02-7.25, 7.52-7.7.63 (2m, **5H,** Ph).

(2S,5R,6R)- **2-Benzyl- 6-tert-butyldiphenylsilyloxy- 5-tert-butyl** diphenylsilyloxymethyl-4-N-methylperhydro-1,4-diazepin-3-one (11). To a solution of **dicyclohexylcarbodiimide** (60.5 mg, 5eq), DMAP.HCI15 (46.5 mg, *5* eq) and DMAP (35.8 mg, 5 eq) in refluxing chloroform (3.35 mL, EtOH free), was added during 2 h a solution of ammonium bis-trifluoroacetate salt 10 (60 mg, 58 µmol) in chloroform (3.35 **mL).** The resulting mixture was maintained at 61 "C for another 2 h and concentrated *in vacuo.* The residue was then dissolved in DMF (1 mL) and both *tert*-butyldiphenylsilyl chloride $(150 \mu L, 10 \text{ eq})$ and imidazole $(79.8 \text{ mg}, 20 \text{ eq})$ were added. After stirring at 45 **"C** for 15 h, ethyl acetate was added to the product mixture which was then hydrolysed by

the addition of a saturated aqueous solution of NaHC03. After decantation and EtOAc extractions ($5x10$ mL), the combined extracts were dried ($Na₂SO₄$) and concentrated *in uacuo.* The crude product absorbed on silica gel was then purified by flash chromatography (cyclohexane:EtOAc:Et3N 7:3:0.03, *Rf 0.54)* to give the expected **perhydro-1,4-diazepin-3-one 11** in 15% yield.16 IH NMR **6** 1.03 *(br.s,* 18H, 2fBu), 2.40 (dd, 1H, $J7.7$ ['] = -15 Hz, $J7.6$ = 9.5 Hz, H-7), 2.96 (s, 3H, NMe), 3.28 (dd, 1H, $J_{CH2Ph} = -13.5 Hz$, $J_{CHPh.2} = 7 Hz$, CHPh), 3.38 (dd, 1H, $J_{CH2Ph} = -13.5 Hz$, $J_{CHPh.2}$ $= 2$ Hz, CH'Ph), 3.43 (dt, 1H, $J_{5,\text{CH'OSi}} = 6.5$ Hz, $J_{5,\text{CH'OSi}} = 6$ Hz, $J_{5.6} = 1.5$ Hz, H-*5),* 3.75 (dd, lH, J7',7 = -15 **Hz,** J7',6 = 7.5 Hz, H-7'), 3.93 (dd, IH, Jcmosi = -1 1 Hz, $J_{CHOSi,5} = 6.5$ Hz, CHOSi), 4.05 (dd and m, 2H, $J_{CH2OSi} = -11$ Hz, $J_{CH7OSi,5} = 6$ Hz, CH'OSi, H-6), 4.93 (br.dd, 1H, $J_{2,\text{CHPh}} = 7$ Hz, $J_{2,\text{CHPh}} = 2$ Hz, H-2), 7.37, 7.59, 7.69 (3m, 25H, Ph); I3C NMR (126 **MHz) 6** 19.0, 26.6, 26.8 (SitBu), 31.9 (NMe), 34.1 (CH2Ph), 48.3 (C-7), 57.1 (C-5), 61.1 (CHzOSi), 62.7 (C-2), 69.8 (C-6), 127.7, 128.2, 129.6, 130.3, 130.4, 132.1, 134.8, 135.2, 135.4, 135.5 (Ph), 170.3 (C-3); MS (70 eV) : **m/z 723 (<1%, M^{o+}-2H-CH3)**, 199 (100%, Ph₂SiOH). The long range ¹H-¹³C correlations observed **(2J** and 3J) are summarized on the following structure :

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REFERENCES and NOTES

- 1. K. Isono, M. Uramoto, H. Kusakabe, K-i. Kimura, K. **Izaki,** C.C. Nelson, and J.A. McCloskey, *J. Antibiof.,* **38,** 1617 (1985).
- K. Kimura, N. Miyata, *G.* Kawanishi, **Y.** Kamio, **K.** Isaki, and K. Isono, *Agric. Biol. Chem.,* **53,** 1811 (1989). *2.*
- **3.** a) M. Ubukata, K. Isono, K-i. Kimura, C.C. Nelson, and J.A. McCloskey, *J. Am. Chem.* **Soc.,llO, 4416 (1988).** b) **M.** Ubukata, K-i. Kimura, K. Isono, C.C. Nelson, J.M. Gregson, and J.A. McCloskey, *J. Org. Chem., 57,* **6392 (1992).**
- **4.** a) M.R. Spada, M. Ubukata, and K. Isono, *Heterocycles,* **34, 1147 (1992).** b) *S.* Knapp, *S.* Nandan, and **L.** Resnick, *Tetrahedron Lett.,* **33,5485 (1992).**
- **5.** I. Charvet, *Thesis*, Université Pierre et Marie Curie, Paris (1995).
- **6.** K.S. Kim, I.H. Cho, Y.H. Ahn, and J.I. Park, *J. Chem. Soc., Perkin Trans. I,* **1783 (1995).**
- **7.** W.J. Moore, and F.A. Luzzio, *Tetrahedron Lett.,* **36,6599 (1995).** The preliminary results described in this communication concern the regiospecific opening of a chiral epoxide by various amino nucleophiles.
- **8.** C. Gravier-Pelletier, J. Dumas, *Y.* Le Merrer, and J.C. Depezay, *J. Carbohydr. Chem.,* **11, 969 (1992)** and references cited therein.
- **9.** a) P. Van de Weghe, and J. Collin, *Tetrahedron Lett.,* **36, 1649 (1995).** b) M. Chini, P. Crotti, L. Favero, F. Macchia, and M. Pineschi, *Tetrahedron Lett., 35,* **433 (1994)** and references cited therein. c) V. Jager, W. Hummer, U. Stahl, and T. Gracza, *Synthesis,* **769 (1991).** d) J. Iqbal, and A. Pandey, *Tetrahedron Lett.,* **31, 575 (1990).** e) M. Fujiwara, M. Imada, A. Baba, and **H.** Matsuda, *Tetrahedron Lett.,* **30, 739 (1989).**
- **10.** a) **H.** Kotsuki, T. Shimanouchi, M. Teraguchi, M. Kataoka, **A.** Tatsukawa, and H.Nishizawa, *Chem. Lett.,* **2159 (1994).** b) T. Tashiro, **S.** Fushiya, and *S.* Nozoe, *Chem. Pharm. Bull.,* **36, 893 (1988).** c) D.F. Burfield, **S.-N.** Gan, and R.H. Smithers, *J. Chem. SOC., Perkin Trans. I,* **666 (1977).**
- 11. In a preliminary study, we observed that during the opening of **7b** by the methyl **N-methyl-L-phenylalaninate,** intramolecular nansesterification reactions occurred.

Such reactions were avoided by N-benzyl protecting the secondary mine on one hand and by using the tert-butyl N-methyl phenylalaninate on the other hand.

No partial epimerization occurred during this sequence of reactions as confirmed by the absence of diastereoisomeric signals in **1H (250 MHz)** and **13C** NMR **(63** MHz) spectra. **12.**

- 13. 8 could also be obtained in 62% yield in presence of ytterbium triflate at 20 °C in two weeks; see 10b.
- This transformation can also be performed by hydrogen in presence of Pd/C 10%, but **required** longer reaction time which led to partial epimerization. 14.
- J. Hu, and M.J. Miller, *Tetrahedron Lett.,* 36, 6379 (1995) and references cited. 15.
- 16. Cyclisation involving a primary amine (RNH2) instead of a secondary one (RNHMe) should occur in a better yield (7 1%) according to reference 6.
- 17. **R.E.** Hurd, and B.K. John, *J. Magn. Reson.,* **91,** 648 (1991).